

# 3<sup>rd</sup> MEETING ON T-CELL AND NK-CELL BASED IMMUNOTHERAPIES FOR LYMPHOID MALIGNANCIES

Jean Lemoine, MD

**Mechanisms of resistance: laboratory evidence**

*Ruella lab, Center for Cellular Immunotherapies, University of Pennsylvania*

BOLOGNA, ROYAL HOTEL CARLTON

September 13-14, 2024



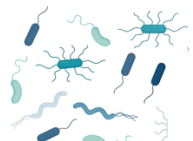
## Disclosures of Jean Lemoine

# Causes of Failure of CART Immunotherapy in the Clinic

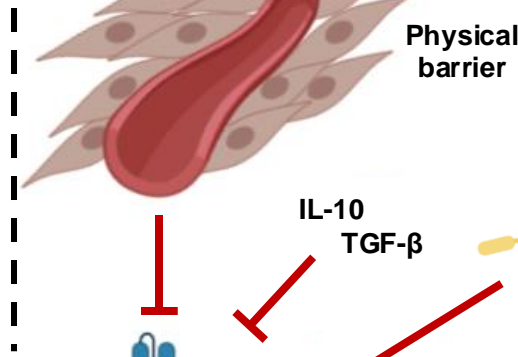
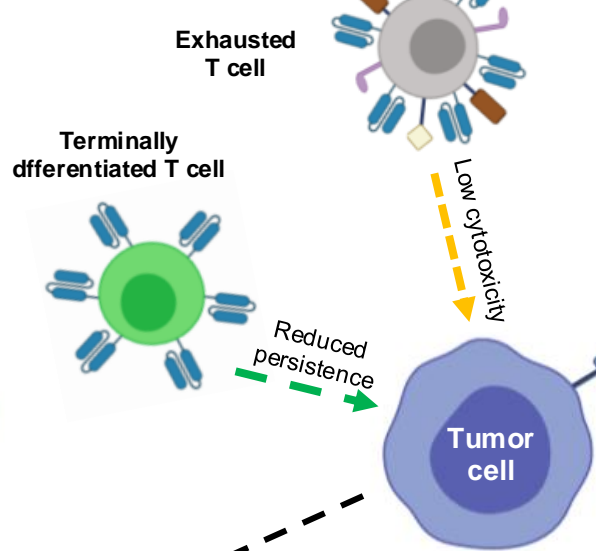
## Pre-infusion barriers

- Low lymphocyte counts
- Progression during manufacturing
- Manufacturing failure
- High Costs
- Access in general

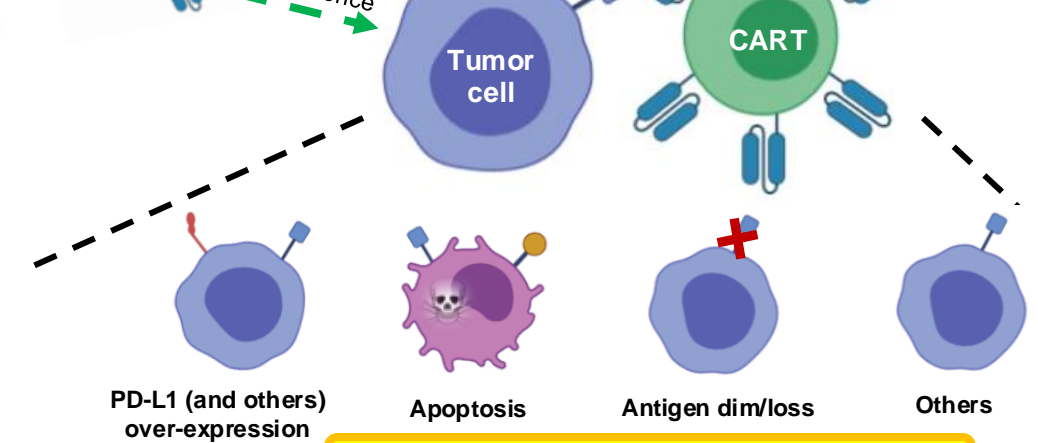
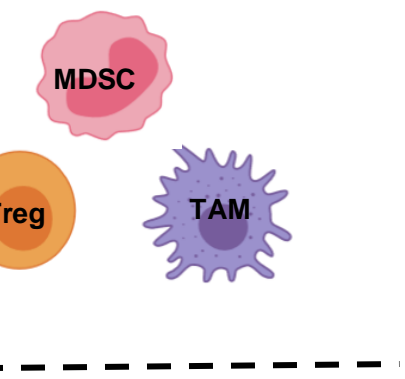
## Host factors

- Tumor burden, previous targeted therapies, etc
  - Immunerejection
  - Microbiota
- 

## CART dysfunction



## Tumor microenvironment



## Tumor-intrinsic mechanisms

- ICANS
- CRS
- Cytopenias
- Off-tumor
- On-target

## Toxicity

- Late neurotoxicity
- Risk of transformation

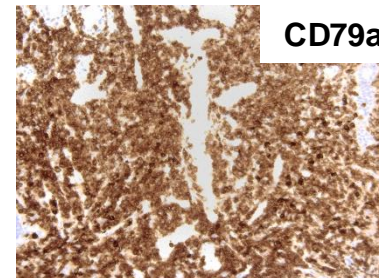
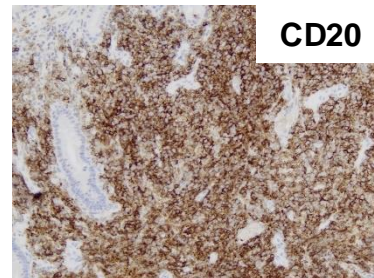
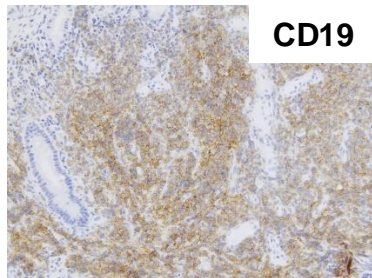
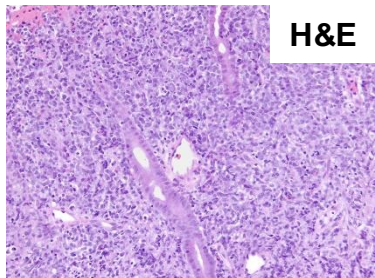
Updated from Ghilardi G., BJH, 2021

# Strategies to overcome CART resistance

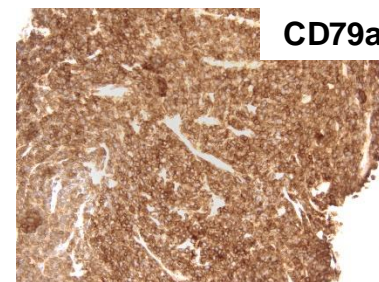
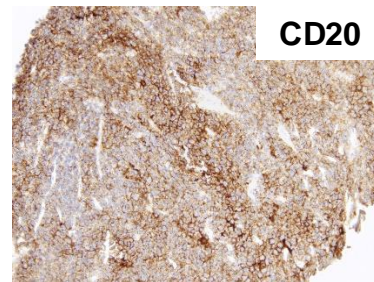
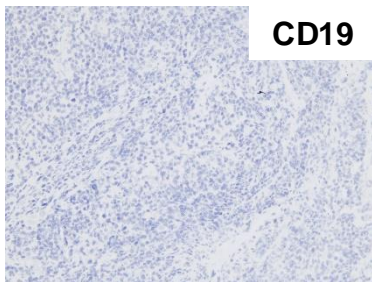
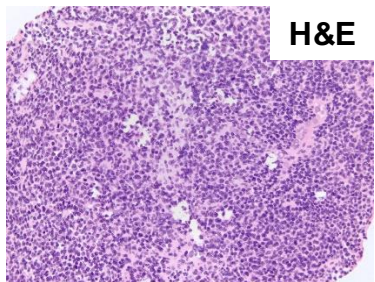
Mechanism of resistance	Potential strategies to overcome	References
Antigen loss or modulation		
Antigen heterogeneity		

# Antigen loss or modulation

Pt #1

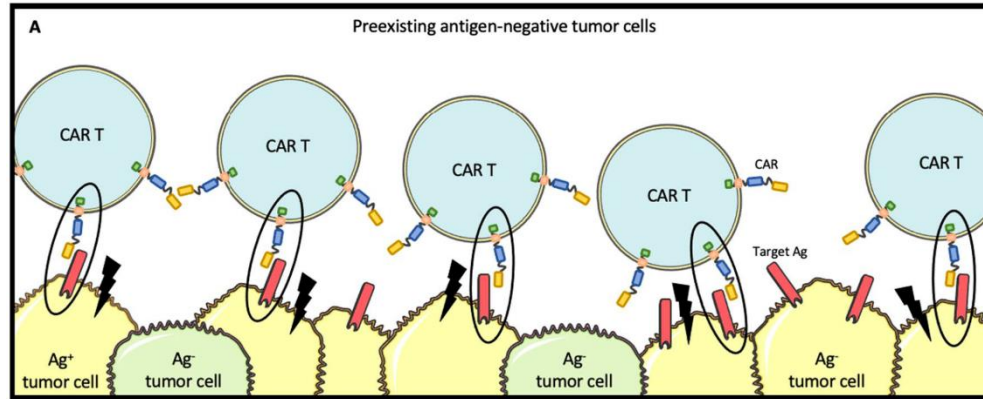


Pt #3



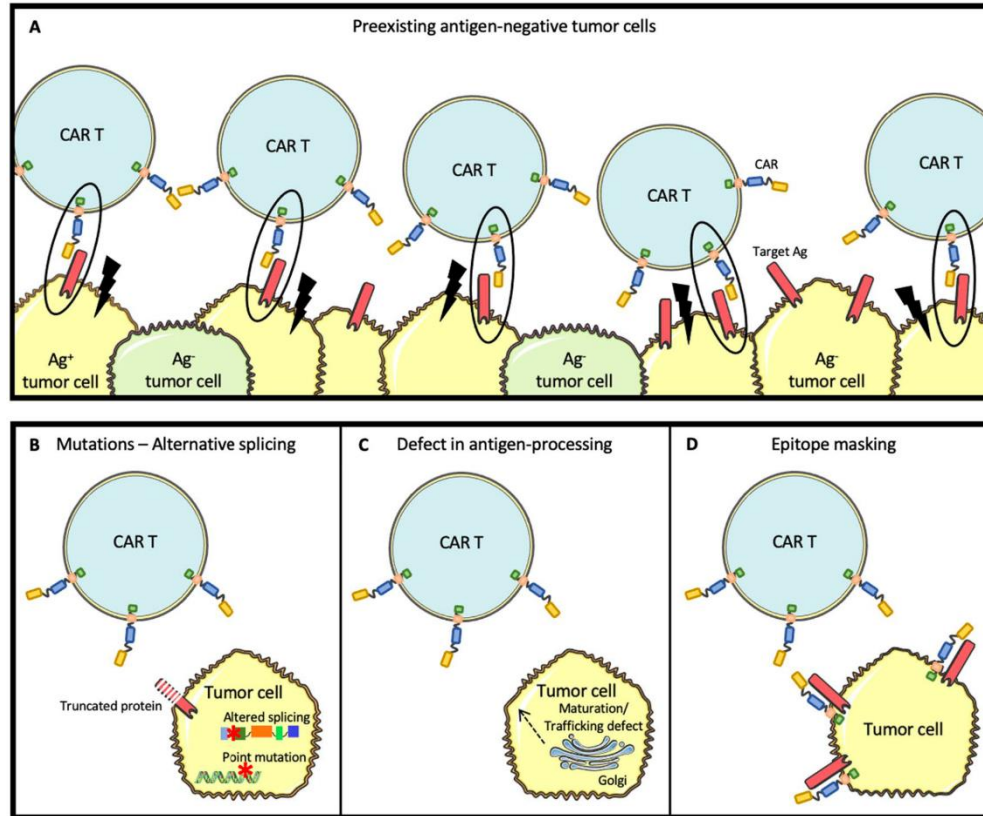
Siddhartha Bhattacharyya

# Mechanisms of Antigen loss or modulation



Lemoine, J., Ruella, M. & Houot, *J Hematol Oncol*, 2021

# Mechanisms of Antigen loss or modulation



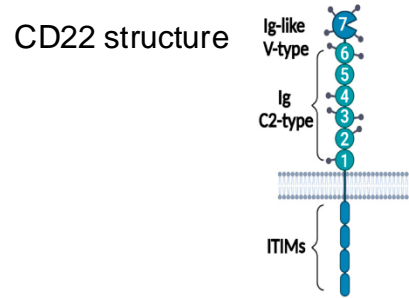
Lemoine, J., Ruella, M. & Houot, *J Hematol Oncol*, 2021

# Strategies to overcome CART resistance

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Antigen heterogeneity		

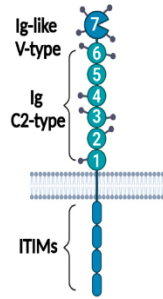


# CD19 and CD22 targeting with CART



# CD19 and CD22 targeting with CART

CD22 structure

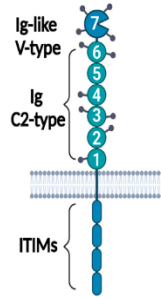


Sample Name	Immunisation	Extracellular Domain(s) of CD22 bound	$K_a$ (1/Ms) $\times 10^4$	$K_d$ (1/s) $\times 10^{-4}$	$K_D$ (nM)
9F9-6	Wistar rats	5 – 6	20.00	2.01	1.00
7E1-2	Wistar rats	5 – 6	10.88	1.25	1.24
9A8-1	Wistar rats	5 – 6	9.57	1.73	1.90
9F8-2	Wistar rats	5 – 6	69.60	87.80	12.60
10C11-6	Wistar rats	5 – 6	12.80	46.70	36.50
9G11-2	Wistar rats	4	2.06	1.57	7.65
8E7-3	Wistar rats	4	2.89	2.56	8.85
7E3-5	Wistar rats	4	2.70	4.53	16.80
10C1-D9	Hyper-Immune mice	3	0.87	0.002	0.03
1G3-4	Wistar rats	3	79.02	3.85	0.52
5H4-9	Hyper-Immune mice	3	4.21	4.57	10.90
4D9-12	Hyper-Immune mice	3	14.60	11.40	8.99

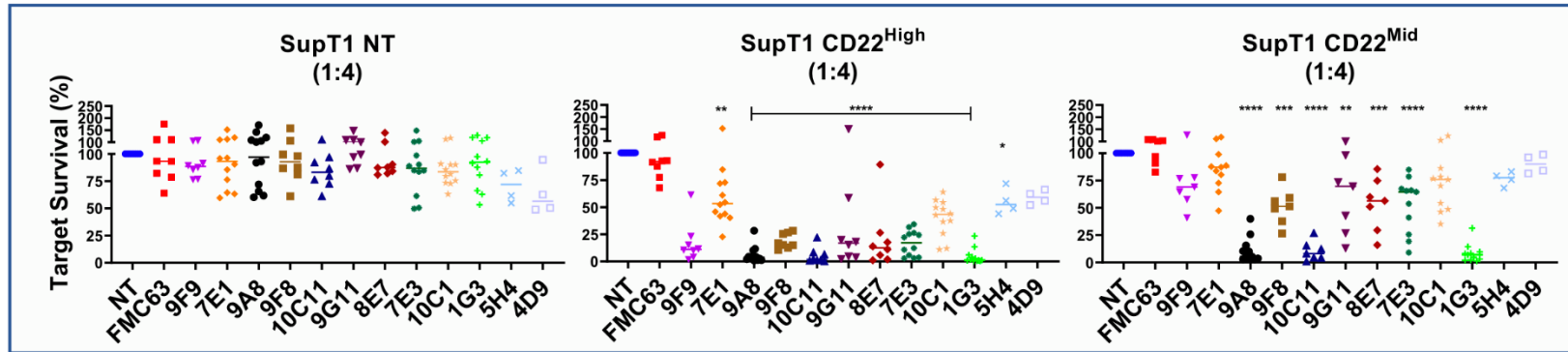
Kokalaki, E. *et al. Mol Ther*, 2023

# CD19 and CD22 targeting with CART

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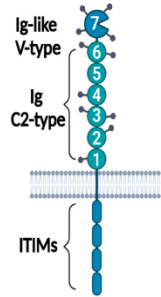
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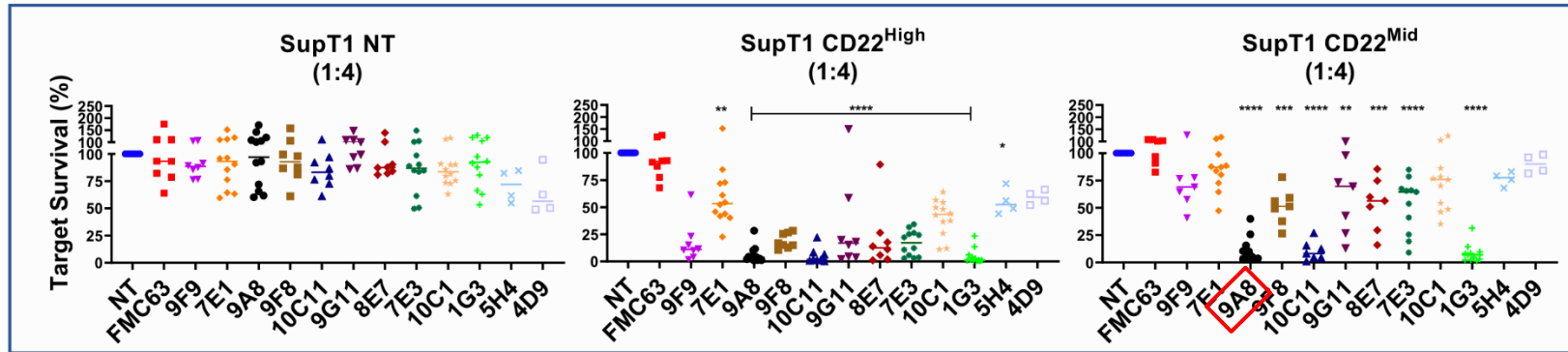
Kokalaki, E. *et al. Mol Ther*, 2023

# CD19 and CD22 targeting with CART

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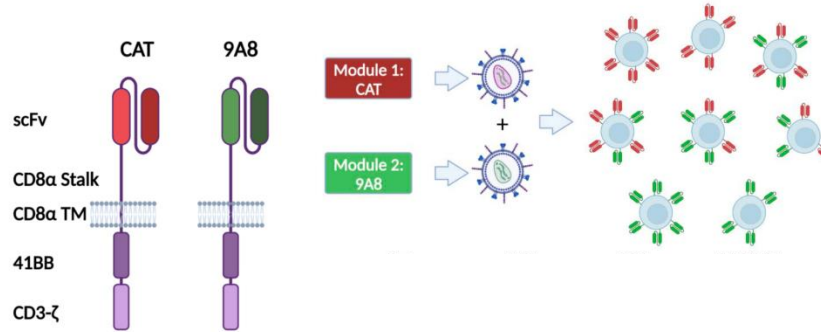


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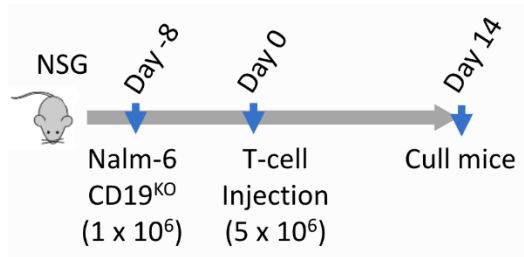
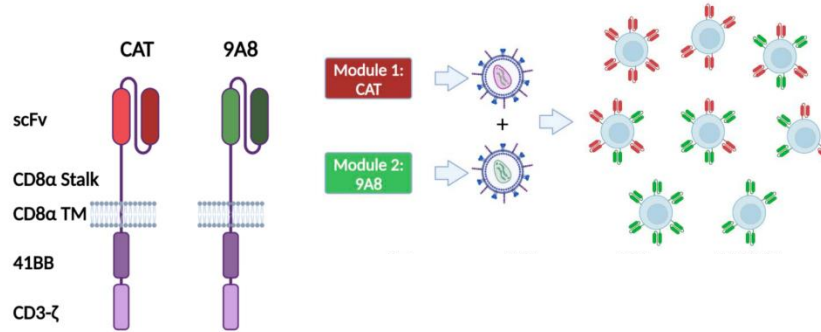


Kokalaki, E. *et al. Mol Ther*, 2023

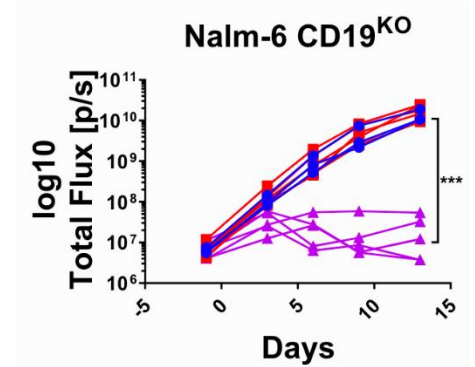
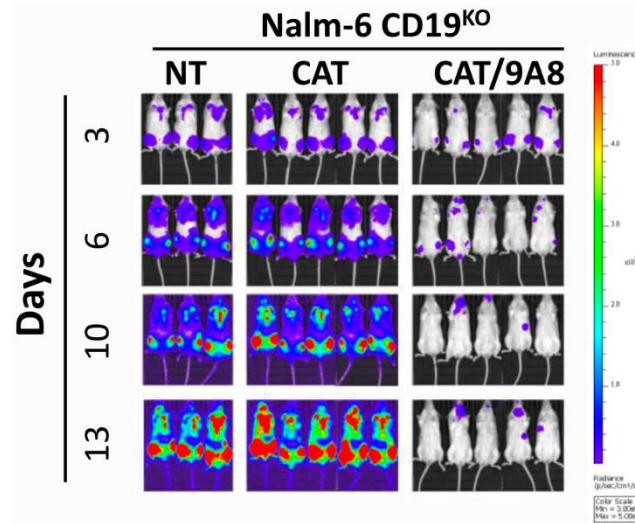
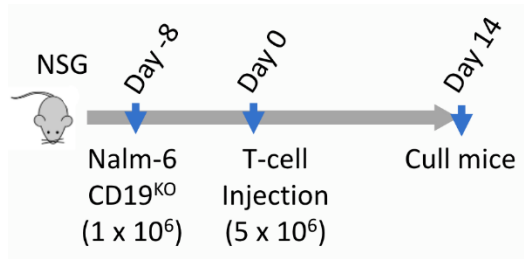
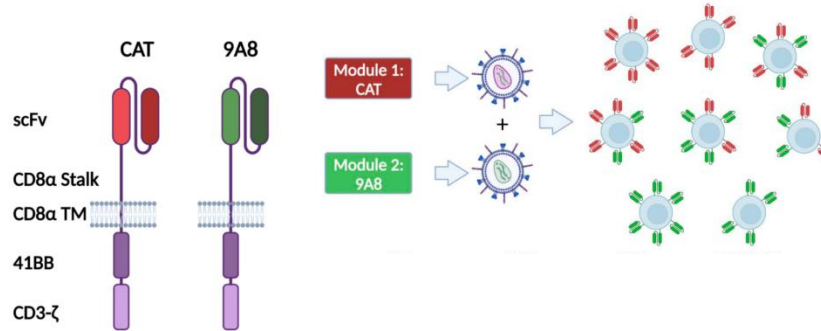
# CD19 and CD22 targeting with CART



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# CD19 and CD22 targeting with CART



Kokalaki, E. et al. *Mol Ther*, 2023

# CD19 and CD22 targeting with CART in the clinic



## CLINICAL TRIALS AND OBSERVATIONS

### CD19/CD22 targeting with cotransduced CAR T cells to prevent antigen-negative relapse after CAR T-cell therapy for B-cell ALL

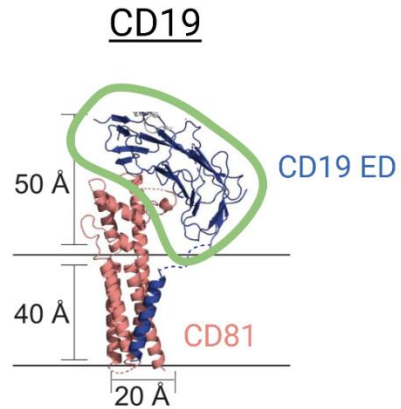
Sara Ghorashian,<sup>1,2,\*</sup> Giovanna Lucchini,<sup>3,\*</sup> Rachel Richardson,<sup>4</sup> Kyvi Nguyen,<sup>4</sup> Craig Terris,<sup>4</sup> Aleks Guvenel,<sup>4</sup> Macarena Oporto-Espuelas,<sup>4</sup> Jenny Yeung,<sup>4</sup> Danielle Pinner,<sup>3</sup> Jan Chu,<sup>3</sup> Lindsey Williams,<sup>3</sup> Ka-Yuk Ko,<sup>3</sup> Chloe Walding,<sup>5</sup> Kelly Watts,<sup>6</sup> Sarah Inglott,<sup>1</sup> Rebecca Thomas,<sup>1</sup> Christopher Connor,<sup>1</sup> Stuart Adams,<sup>1</sup> Emma Gravett,<sup>1</sup> Kimberly Gilmour,<sup>7</sup> Alka Lal,<sup>8</sup> Sangeetha Kunaseelan,<sup>8</sup> Bilyana Popova,<sup>8</sup> Andre Lopes,<sup>8</sup> Yenting Ngai,<sup>8</sup> Allan Hackshaw,<sup>8</sup> Evangelia Kokalaki,<sup>9</sup> Milena Balasch Carulla,<sup>3</sup> Khushnuma Mullanfiroze,<sup>3</sup> Arina Lazareva,<sup>3</sup> Vesna Pavasovic,<sup>1</sup> Anupama Rao,<sup>1</sup> Jack Bartram,<sup>1</sup> Ajay Vora,<sup>1</sup> Robert Chiesa,<sup>3</sup> Juliana Silva,<sup>3</sup> Kanchan Rao,<sup>4</sup> Denise Bonney,<sup>6</sup> Robert Wynn,<sup>6</sup> Martin Pule,<sup>9</sup> Rachael Hough,<sup>5</sup> and Persis J. Amrolia<sup>3,4</sup>



# Strategies to overcome CART resistance

Mechanism of resistance	Potential strategies to overcome	References
Antigen loss or modulation	<ul style="list-style-type: none"> <li>- Develop CART targeting multiple antigens</li> </ul>	<ul style="list-style-type: none"> <li>- Kokalaki, E. <i>et al.</i> Dual targeting of CD19 and CD22 against B-ALL using a novel high-sensitivity aCD22 CAR. <i>Mol Ther</i>, (2023)</li> </ul>
Antigen heterogeneity	<ul style="list-style-type: none"> <li>- Develop CAR with high sensitivity for the antigen</li>   <li>- Use CAR targeting epitopes less prone to mutation</li> </ul>	<ul style="list-style-type: none"> <li>- Chu, F. <i>et al.</i> Chimeric antigen receptor T cells to target CD79b in B-cell lymphomas. <i>J Immunother Cancer</i>, (2023)</li> <li>- Zhang, Y. <i>et al.</i> Safety and efficacy of a novel anti-CD19 chimeric antigen receptor T cell product targeting a membrane-proximal domain of CD19 with fast on- and off-rates against non-Hodgkin lymphoma: a first-in-human study. <i>Mol Cancer</i>, (2023)</li>   <li>- Cohen, IJ. <i>et al.</i> Chimeric Antigen Receptor T Cells (CART) against the IGHV4-34 B cell Receptor Eliminate Neoplastic B Cells and Reduce Antigen-Negative Escape while Sparing the 2 Healthy B cell Repertoire. Submitted</li> </ul>

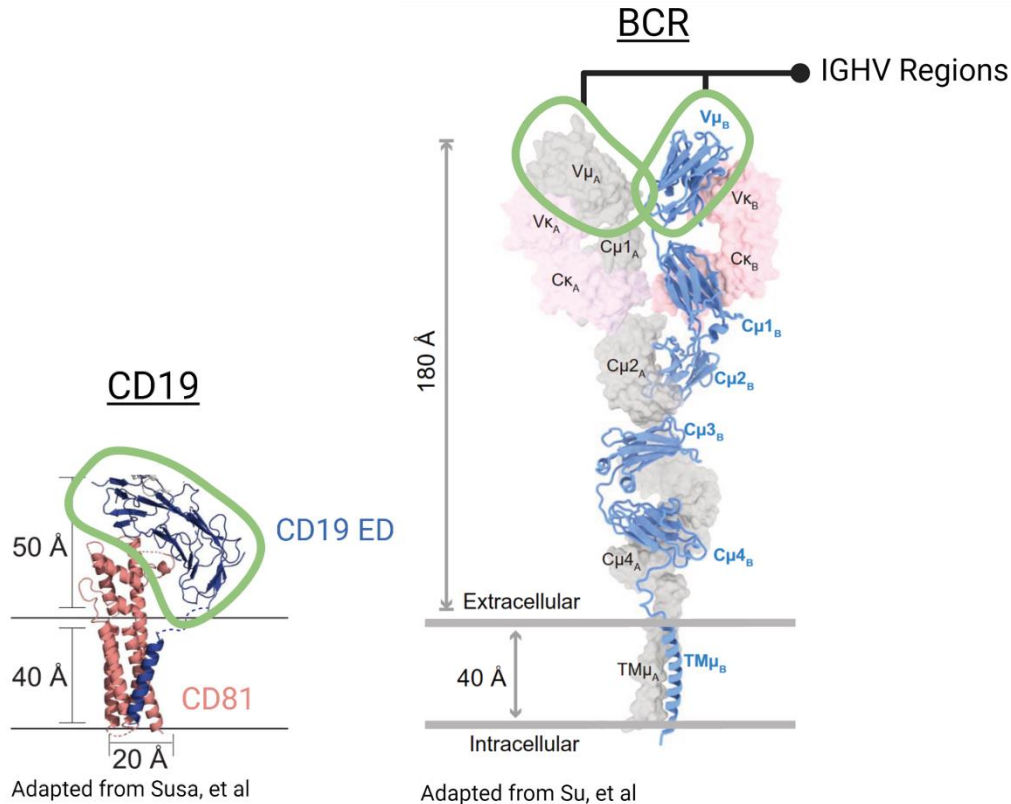
# CART targeting epitopes less prone to mutation



Adapted from Susa, et al

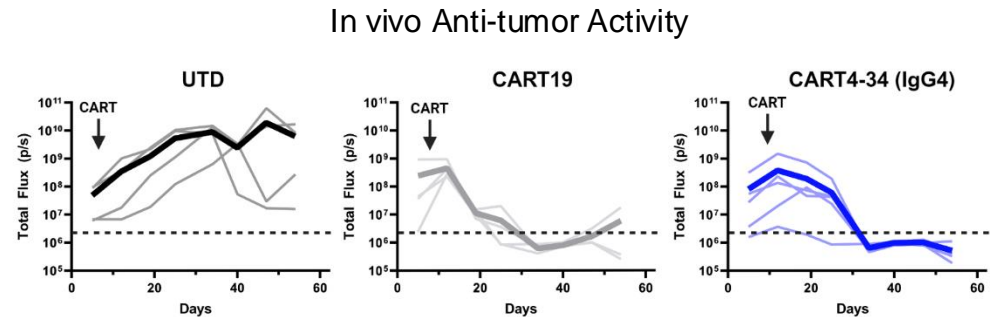
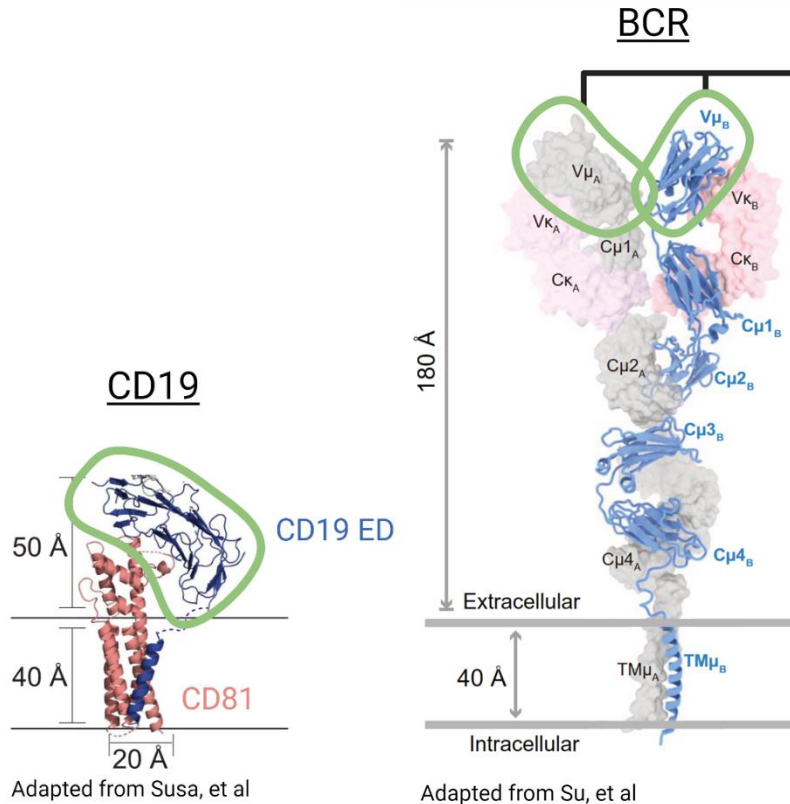
Cohen IJ *et al.* Submitted  
Susa, K *et al.* Science 2021  
Su, Q *et al.* Science, 2022

# CART targeting epitopes less prone to mutation



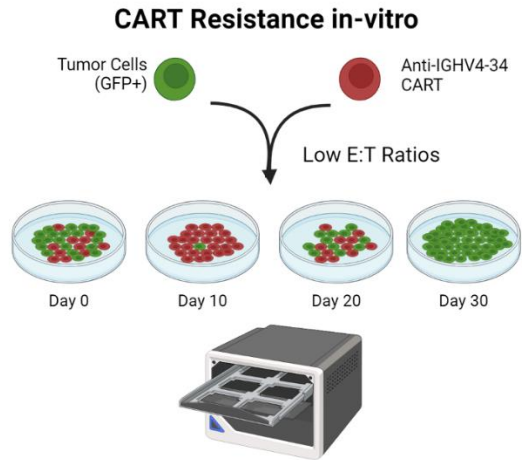
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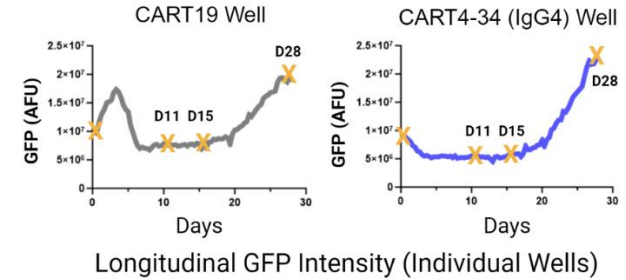
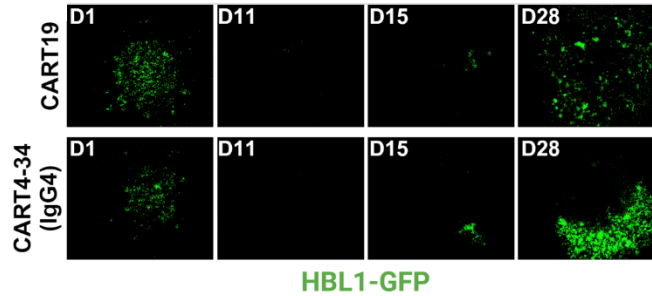
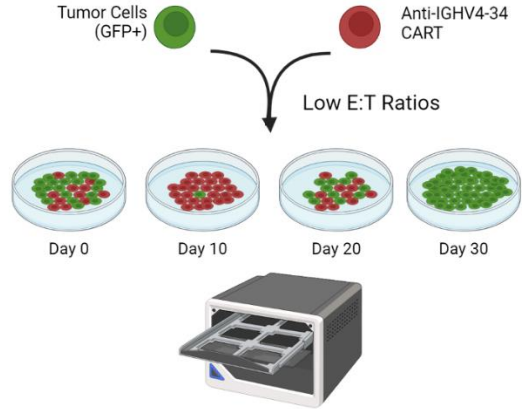
# CART targeting epitopes less prone to mutation



Long-term 96-well plate fluorescence imaging

# CART targeting epitopes less prone to mutation

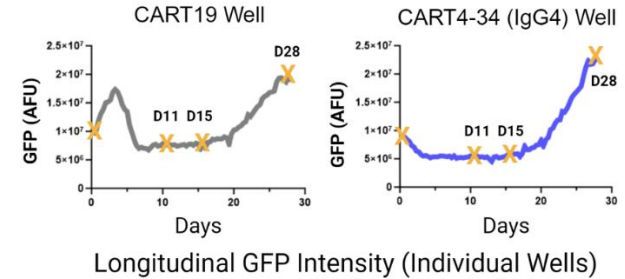
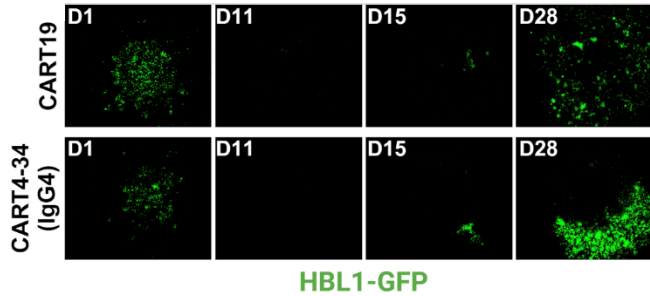
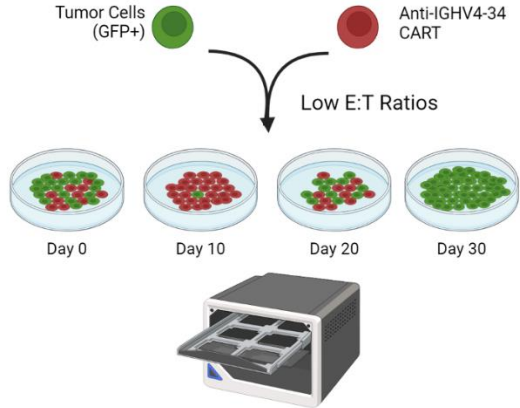
## CART Resistance in-vitro



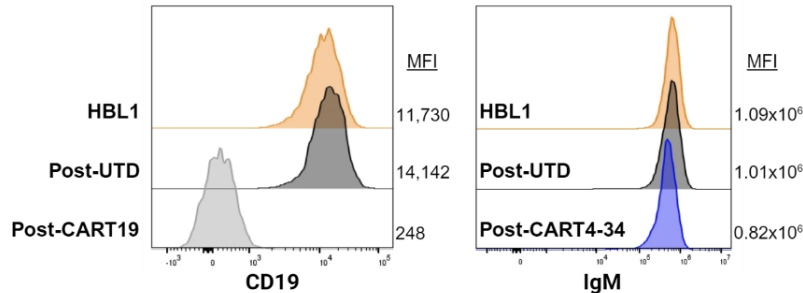
Cohen IJ *et al.* submitted

# CART targeting epitopes less prone to mutation

## CART Resistance in-vitro



## Antigen Expression Post-CART



Cohen IJ *et al.* submitted

# Strategies to overcome CART resistance

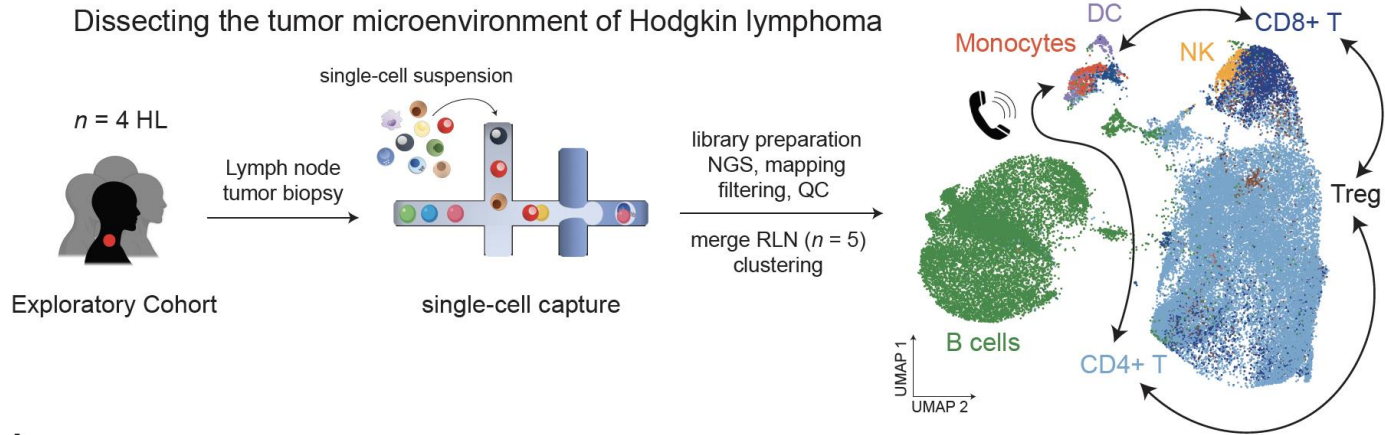
Mechanism of resistance	Potential strategies to overcome	References
CART exhaustion		
Lack of costimulatory signals		
Immunosuppressive microenvironment		



# Strategies to overcome CART resistance

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CART exhaustion	- CART gene editing for improved activation	- Guruprasad, P. <i>et al.</i> The BTLA-HVEM axis restricts CAR T cell efficacy in cancer. <i>Nat Immunol</i> , (2024)
Lack of costimulatory signals	- Optimize co-stimulatory domains	- Patel, R. P. <i>et al.</i> CD5 deletion enhances the antitumor activity of adoptive T cell therapies. <i>Sci Immunol</i> , (2024)
Immunosuppressive microenvironment		- Doan, A. E. <i>et al.</i> FOXO1 is a master regulator of memory programming in CAR T cells. <i>Nature</i> , (2024)

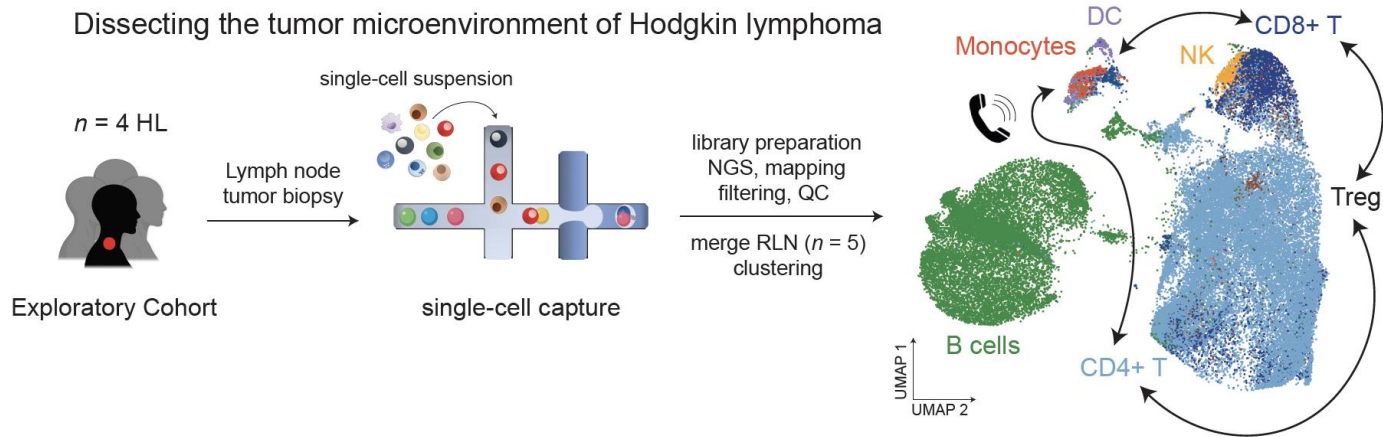
# The BTLA-HVEM axis is enriched in Hodgkin Lymphoma



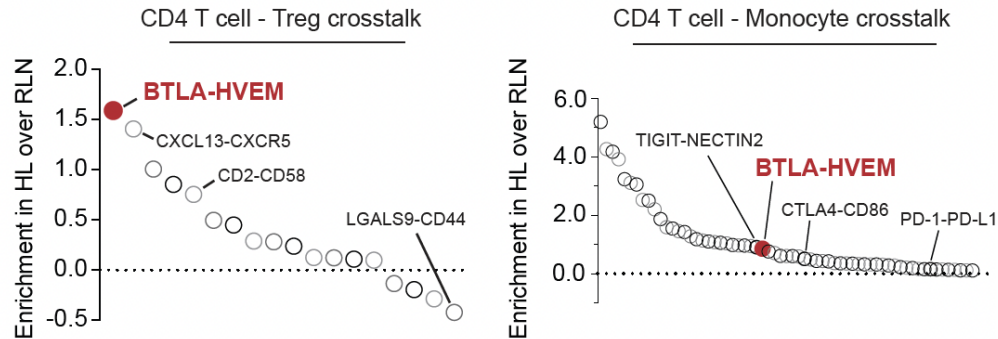
Guruprasad, P. *et al. Nat Immunol* (2024)

# The BTLA-HVEM axis is enriched in Hodgkin Lymphoma

Dissecting the tumor microenvironment of Hodgkin lymphoma

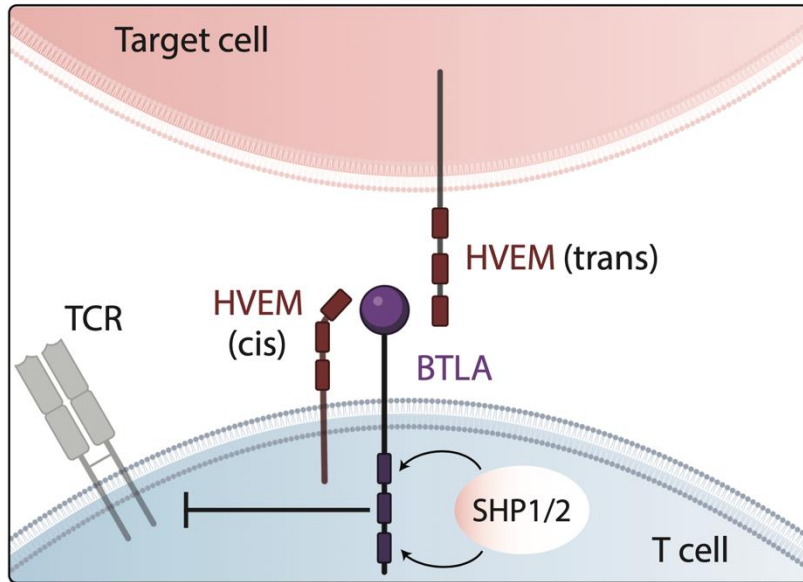


Ligand-receptor interactions between T and immunosuppressive cells



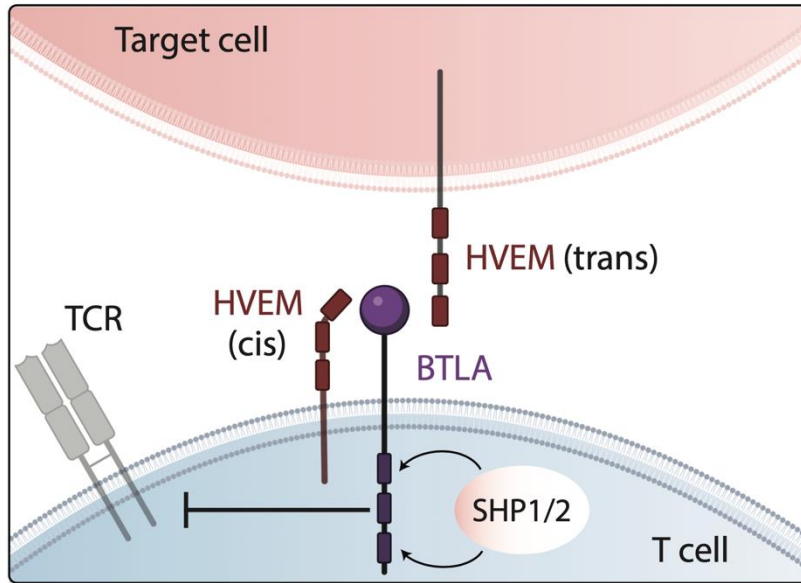
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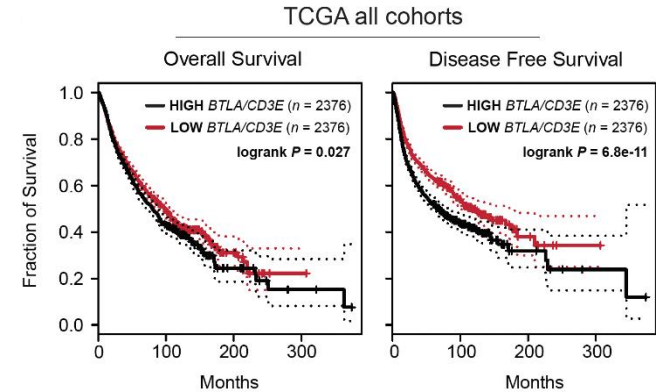


Guruprasad, P. *et al. Nat Immunol* (2024)

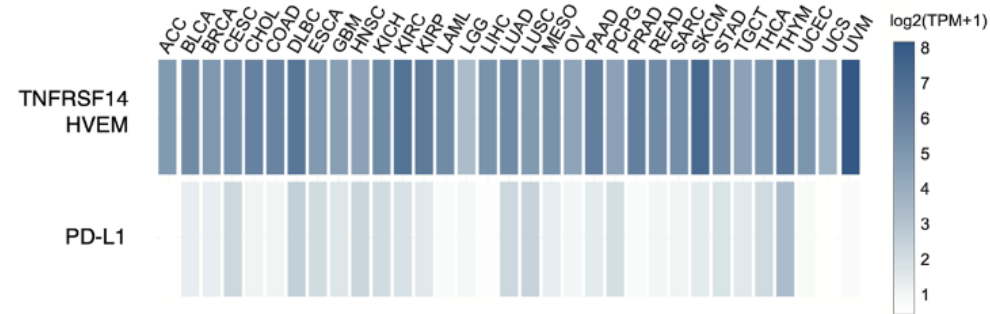
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## BTLA expression in TCGA

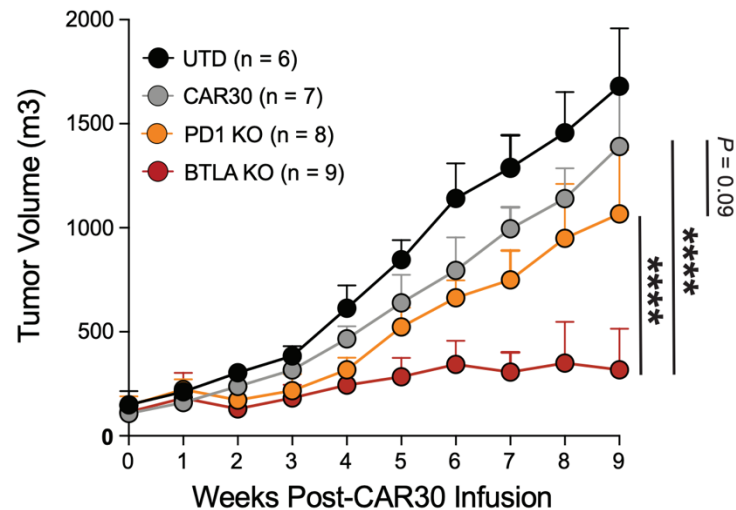
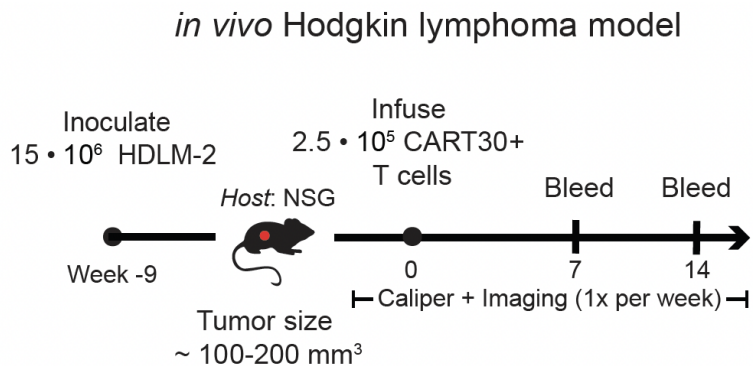


## HVEM expression in TCGA



Guruprasad, P. *et al. Nat Immunol* (2024)

# BTLA KO compared to PD-1 KO in CART30 for Hodgkin Lymphoma



Guruprasad, P. *et al. Nat Immunol* (2024)

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Lack of costimulatory signals		
Immunosuppressive microenvironment	<ul style="list-style-type: none"> <li>- Combination with immune checkpoint blockers</li> </ul>	<ul style="list-style-type: none"> <li>- Rupp, L. J. <i>et al.</i> CRISPR/Cas9-mediated PD-1 disruption enhances anti-tumor efficacy of human chimeric antigen receptor T cells. <i>Sci Rep</i>, (2017)</li> <li>- Pérez-Moreno, M. A. <i>et al.</i> Combined or Sequential Treatment with Immune Checkpoint Inhibitors and Car-T Cell Therapies for the Management of Haematological Malignancies: A Systematic Review. <i>Int J Mol Sci</i>, (2023)</li> </ul>

# Combination with checkpoint blockers

Author, Year	Pathology	Study Treatment	Patients (N)	Response N (%)
Li et al. [28], 2017	B-ALL	Anti-CD19 + Nivolumab or Pembrolizumab	14	ORR: 6 (42.9%) - CR: 2 (14.3%) - PR: 4 (28.6%) PD: 1 (7.1%)
Maude et al. [29], 2017	B-ALL	Tisa-cel+ Pembrolizumab	4	ORR: 2 (50.0%) - CR: 1 (25.0%) - PR: 1 (25.0%)
Cao et al. [30], 2019	B cell-NHL	Anti-CD19+ Nivolumab	11	ORR: 9 (81.8%) - CR: 5 (45.5%) - PR: 4 (36.4%)
Siddiqi et al. [31], 2019 (PLATFORM study)	B cell-NHL	Liso-cel + Durvalumab	18 (11 *)	ORR: 10 (90.9%) - CR: 7 (63.6)
Jacobson et al. [33], 2020 (ZUMA-6 trial)	B cell-NHL	Axi-cel + Atezolizumab	28	ORR: 21 (75.0%) - CR: 13 (46.4%)
Ramakrishnan et al. [34], 2020 (ALEXANDER trial)	B cell-NHL	AUTO3 + Pembrolizumab	33 (29 *)	ORR: 20 (69.0%) - CR: 15 (51.7%)
Chong et al. [35], 2022	B cell-NHL	Tisa-cel + Pembrolizumab	12	ORR: 3 (25.0%) - CR: 1 (8.3%) - PR: 2 (16.7%) PD: 8 (66.7%) SD: 1 (8.3%)
Hirayama et al. [36], 2022	B cell-NHL	Anti-CD19+ Durvalumab	29 (26 *)	ORR: 9 (34.6%) - RC: 7 (26.9%)
Jaeger et al. [37], 2023 (PORTIA trial)	B cell-NHL	Tisa-cel + Pembrolizumab	12	ORR: 6 (50.0%) - CR: 4 (33.3%) - PR: 2 (16.7%) PD: 6 (50.0%)

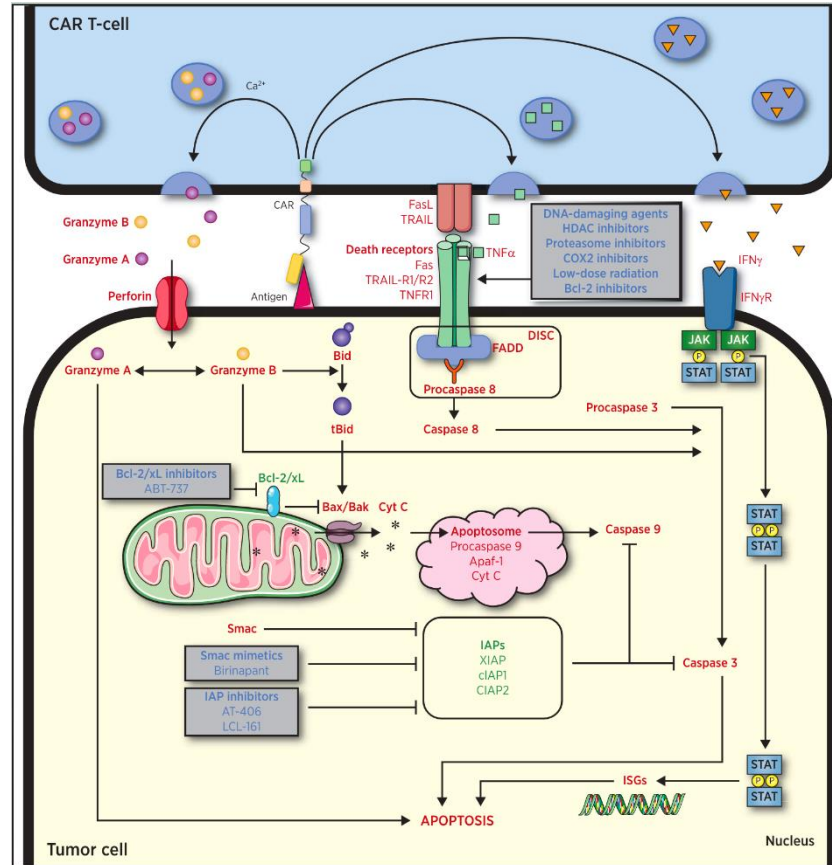
Pérez-Moreno, M. A. *et al. Int J Mol Sci*, (2023)



# Strategies to overcome CART resistance

Mechanism of resistance	Potential strategies to overcome	References
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Lack of costimulatory signals	- Optimize co-stimulatory domains	- Patel, R. P. <i>et al.</i> CD5 deletion enhances the antitumor activity of adoptive T cell therapies. <i>Sci Immunol</i> , (2024)
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	- Combination with small molecules	- Rupp, L. J. <i>et al.</i> CRISPR/Cas9-mediated PD-1 disruption enhances anti-tumor efficacy of human chimeric antigen receptor T cells. <i>Sci Rep</i> , (2017) - Pérez-Moreno, M. A. <i>et al.</i> Combined or Sequential Treatment with Immune Checkpoint Inhibitors and Car-T Cell Therapies for the Management of Haematological Malignancies: A Systematic Review. <i>Int J Mol Sci</i> , (2023) - Lemoine, J., Ruella, M. & Houot, R. Overcoming intrinsic resistance of cancer cells to CAR T-cell killing. <i>Clin Cancer Res</i> , (2021) - Michie, J. <i>et al.</i> Antagonism of IAPs Enhances CAR T-cell Efficacy. <i>Cancer Immunology Research</i> , (2019)

# Combination with pro-apoptotic small molecules



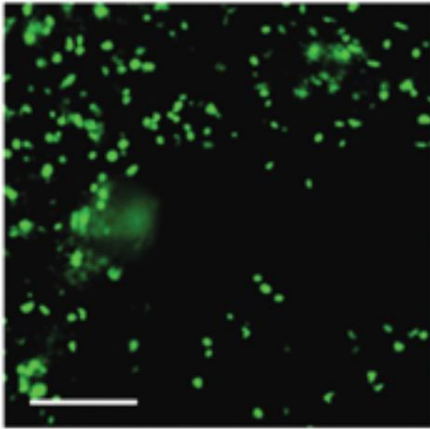
Lemoine, J., Ruella, M. & Houot, R. *Clin Cancer Res*, (2021)

# Strategies to overcome CART resistance

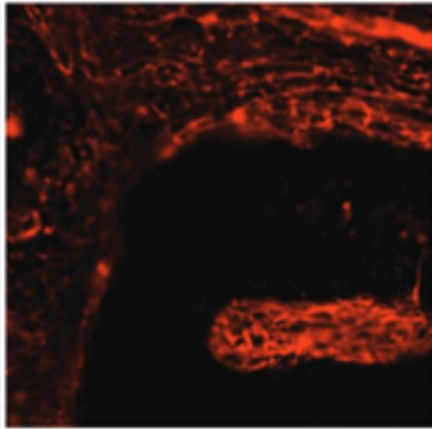
Mechanism of resistance	Potential strategies to overcome	References
Poor tumor infiltration		

# Poor tumor infiltration

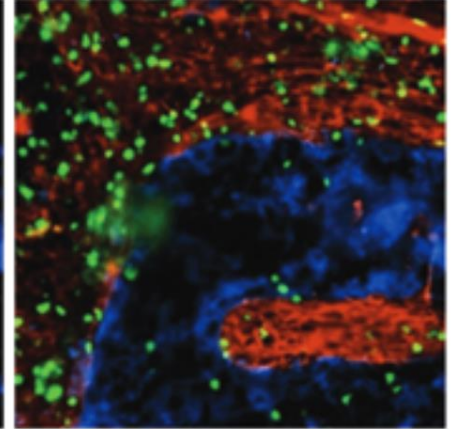
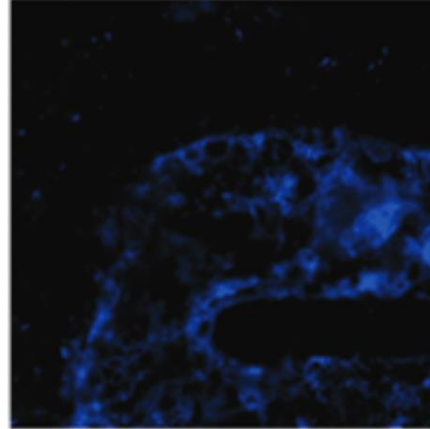
T cells



Fibronectin



EpCAM



Salmon, H. *et al. J Clin Invest*, (2012)

# Strategies to overcome CART resistance

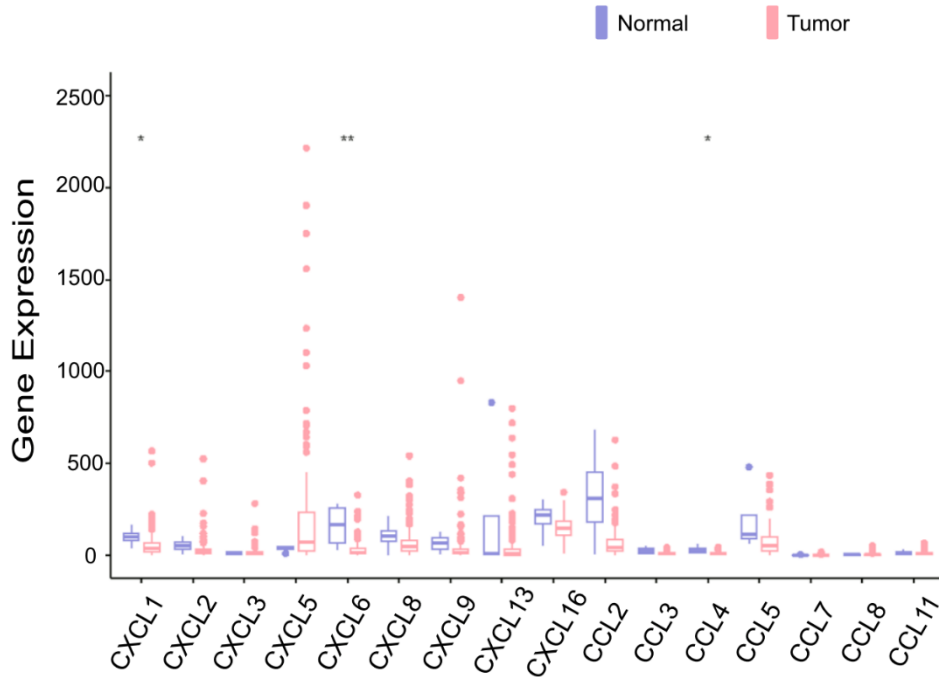
Mechanism of resistance	Potential strategies to overcome	References
Poor tumor infiltration	- Local delivery	- Adusumilli, P. S. <i>et al.</i> Regional delivery of mesothelin-targeted CAR T cell therapy generates potent and long-lasting CD4-dependent tumor immunity. <i>Sci Transl Med</i> , (2014) - Brown, C. E. <i>et al.</i> Optimization of IL13R $\alpha$ 2-Targeted Chimeric Antigen Receptor T Cells for Improved Anti-tumor Efficacy against Glioblastoma. <i>Mol Ther</i> , (2018)

# Strategies to overcome CART resistance

Mechanism of resistance	Potential strategies to overcome	References
Poor tumor infiltration	<ul style="list-style-type: none"><li>- Local delivery</li><li>- Enhancing tumor infiltration through chemokine axis</li></ul>	<ul style="list-style-type: none"><li>- Adusumilli, P. S. <i>et al.</i> Regional delivery of mesothelin-targeted CAR T cell therapy generates potent and long-lasting CD4-dependent tumor immunity. <i>Sci Transl Med</i>, (2014)</li><li>- Brown, C. E. <i>et al.</i> Optimization of IL13R<math>\alpha</math>2-Targeted Chimeric Antigen Receptor T Cells for Improved Anti-tumor Efficacy against Glioblastoma. <i>Mol Ther</i>, (2018)</li><li>- Dai, Z. <i>et al.</i> Ectopic CXCR2 expression cells improve the anti-tumor efficiency of CAR-T cells and remodel the immune microenvironment of pancreatic ductal adenocarcinoma. <i>Cancer Immunol Immunother</i>, (2024)</li></ul>

# CXCR2 expressing CAR to enhance infiltration in pancreatic adenocarcinoma

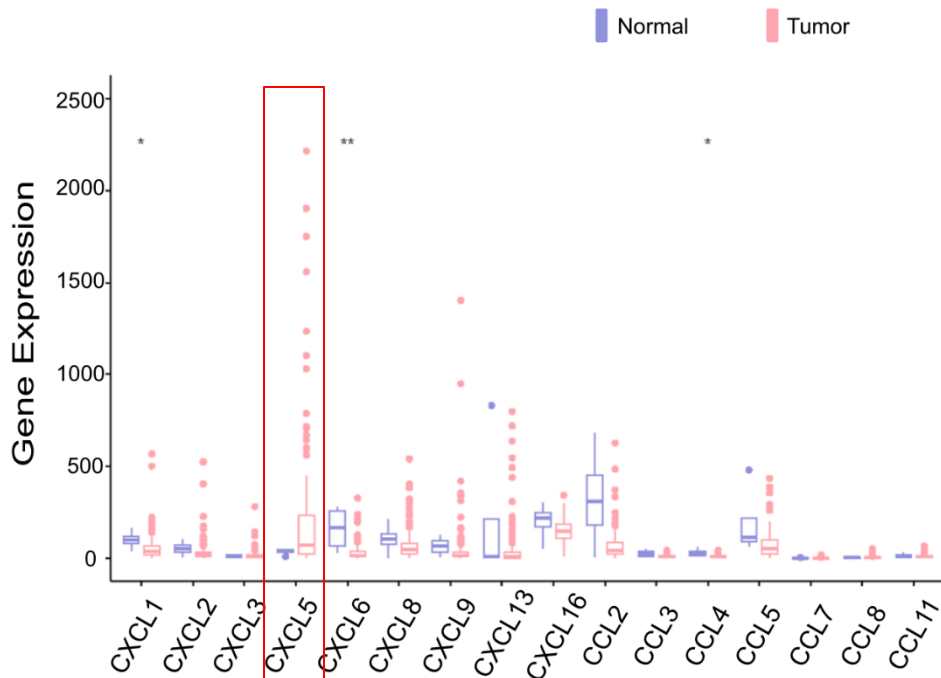
Differential expression of chemokines



Proudfoot, A. E. I. *Nat Rev Immunol*, (2002)  
Dai, Z. *et al. Cancer Immunol Immunother*, (2024)

# CXCR2 expressing CAR to enhance infiltration in pancreatic adenocarcinoma

Differential expression of chemokines

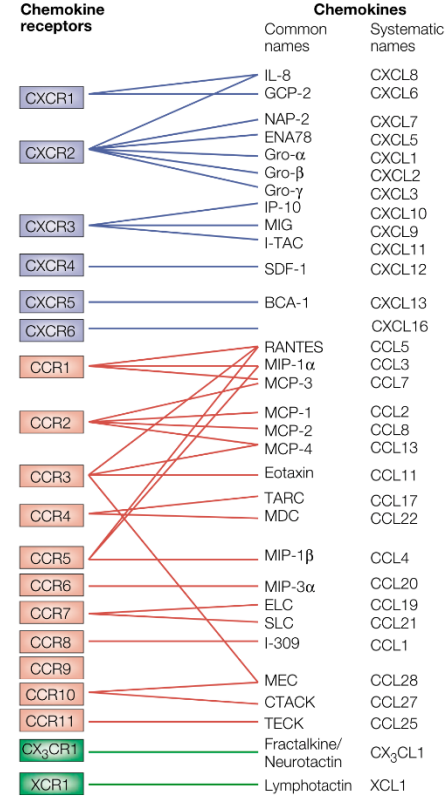
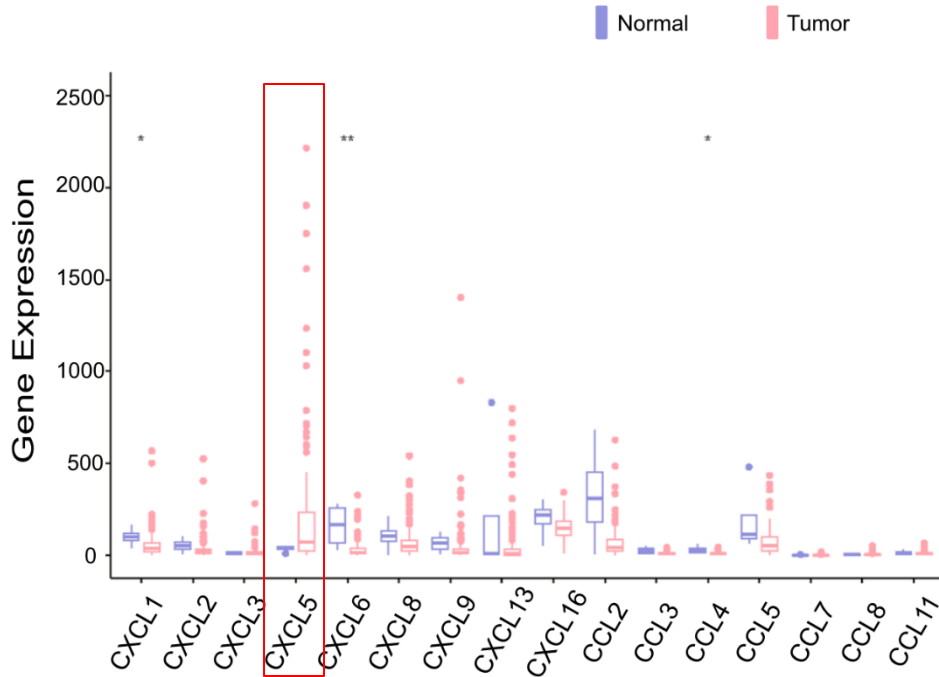


Proudfoot, A. E. I. *Nat Rev Immunol*, (2002)  
Dai, Z. *et al. Cancer Immunol Immunother*, (2024)



# CXCR2 expressing CAR to enhance infiltration in pancreatic adenocarcinoma

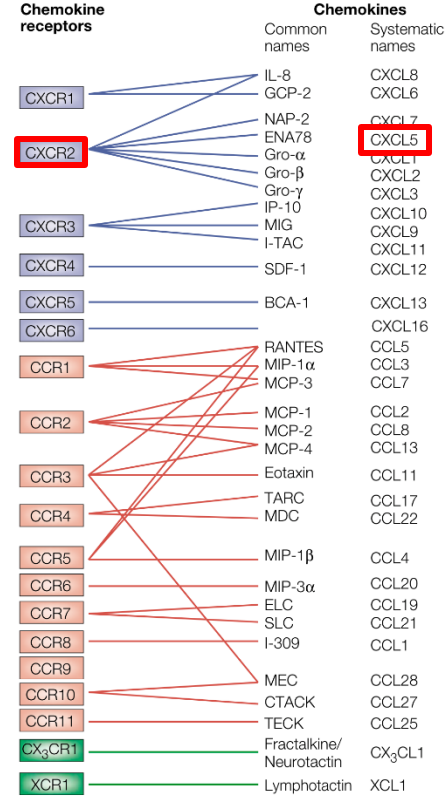
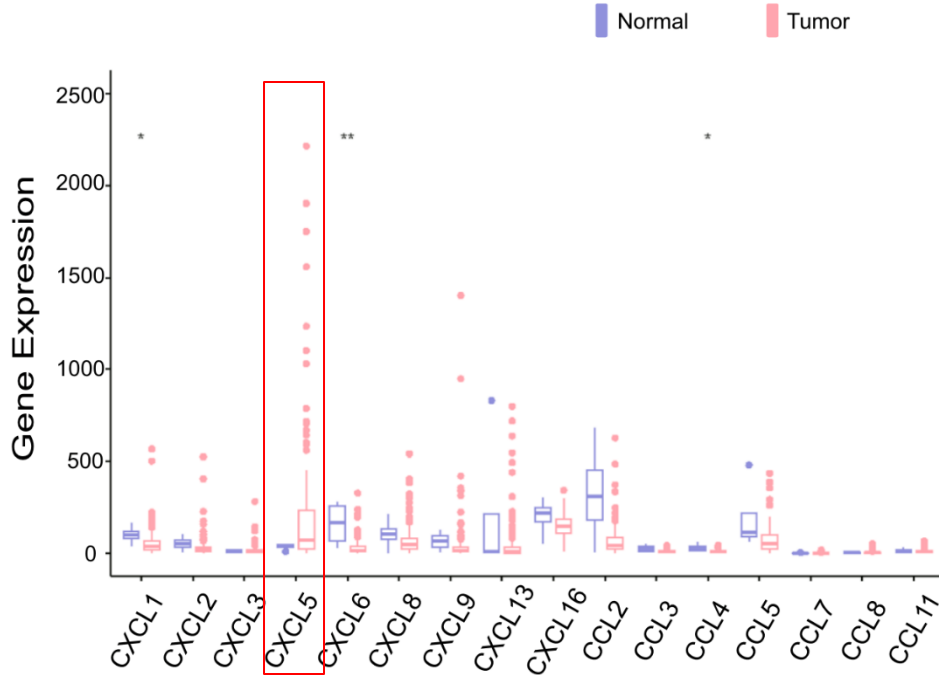
Differential expression of chemokines



Proudfoot, A. E. I. *Nat Rev Immunol*, (2002)  
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Differential expression of chemokines



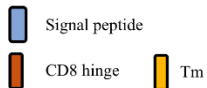
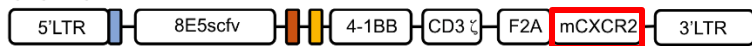
Proudfoot, A. E. I. *Nat Rev Immunol*, (2002)  
 Dai, Z. *et al. Cancer Immunol Immunother*, (2024)

# CXCR2 expressing CAR to enhance infiltration in pancreatic adenocarcinoma

Claudin18.2 CAR



CXCR2 CAR



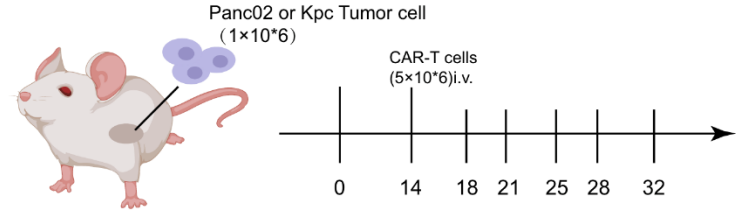
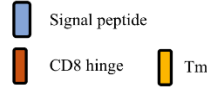
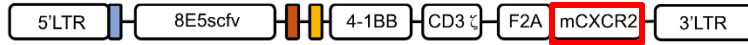
Dai, Z. *et al. Cancer Immunol Immunother*, (2024)

# CXCR2 expressing CAR to enhance infiltration in pancreatic adenocarcinoma

Claudin18.2 CAR

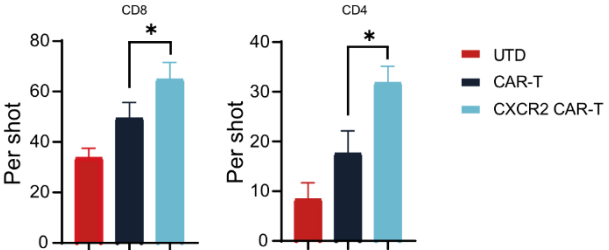
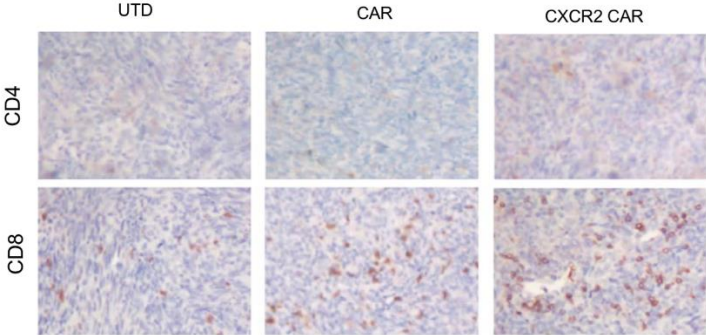
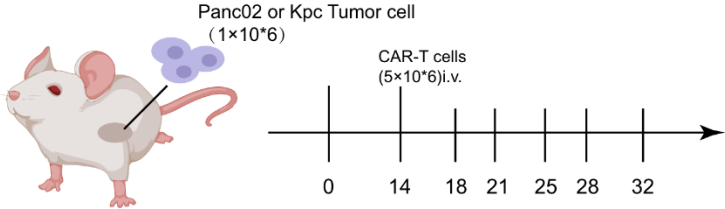
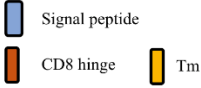


CXCR2 CAR



Dai, Z. *et al. Cancer Immunol Immunother.* (2024)

# CXCR2 expressing CAR to enhance infiltration in pancreatic adenocarcinoma



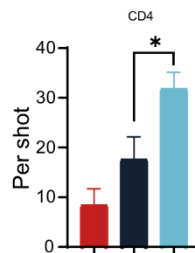
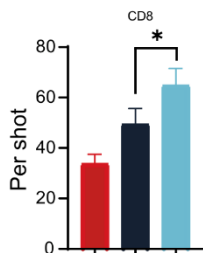
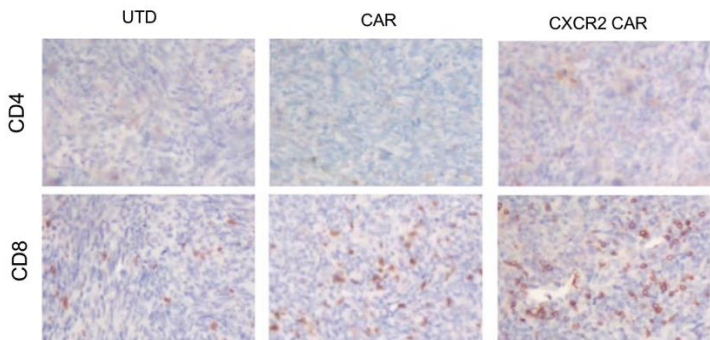
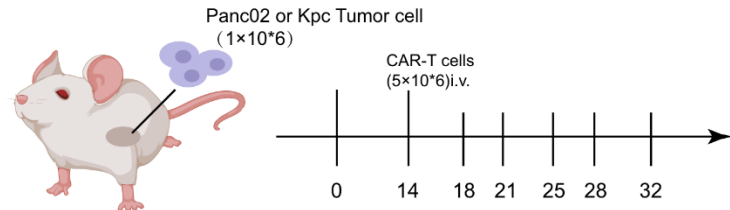
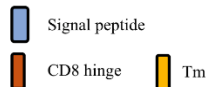
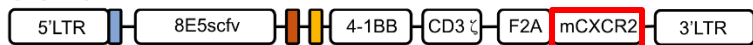
Dai, Z. et al. *Cancer Immunol Immunother.* (2024)

# CXCR2 expressing CAR to enhance infiltration in pancreatic adenocarcinoma

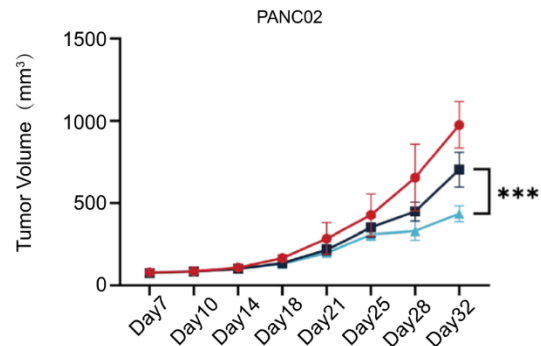
Claudin18.2 CAR



CXCR2 CAR



Legend for tumor volume graph: Red = UTD, Black = CAR-T, Blue = CXCR2 CAR-T

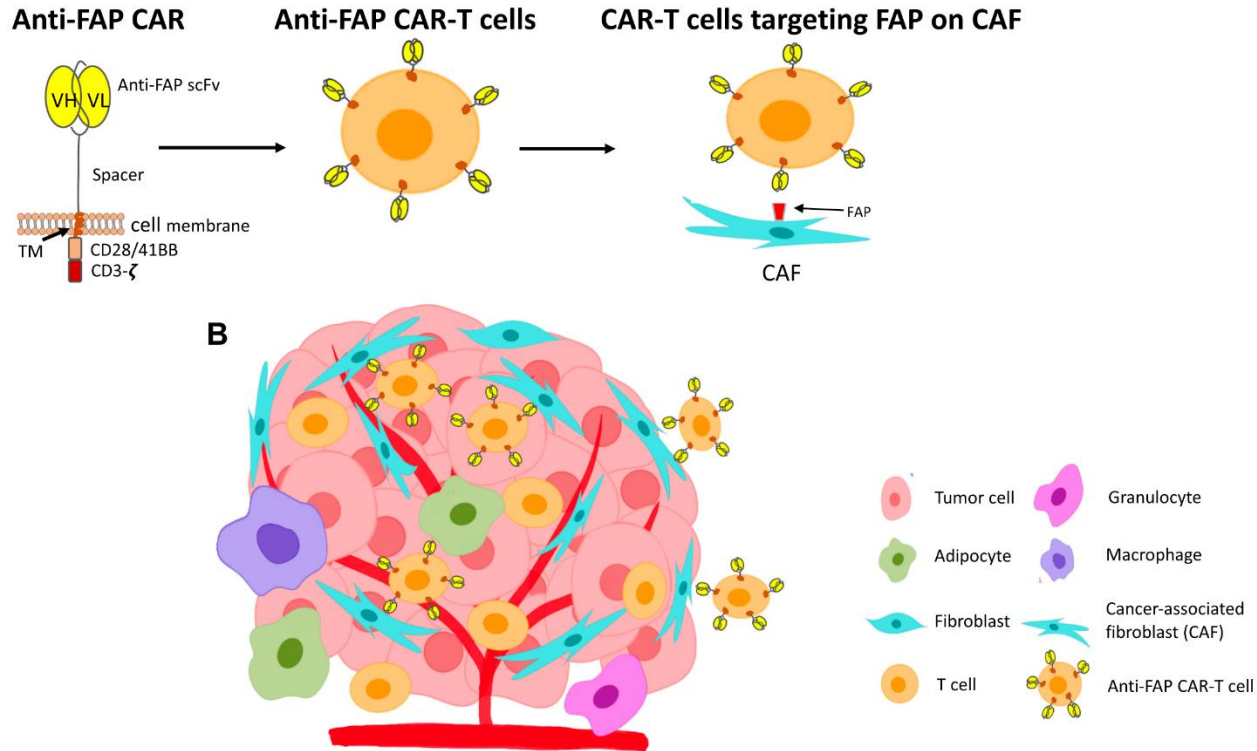


Dai, Z. *et al. Cancer Immunol Immunother.* (2024)

# Strategies to overcome CART resistance

Mechanism of resistance	Potential strategies to overcome	References
Poor tumor infiltration	<ul style="list-style-type: none"> <li>- Local delivery</li>   <li>- Enhancing tumor infiltration through chemokine axis</li>   <li>- Enhancing tumor infiltration targeting extra-cellular matrix fibers or fibroblasts</li> </ul>	<ul style="list-style-type: none"> <li>- Adusumilli, P. S. <i>et al.</i> Regional delivery of mesothelin-targeted CAR T cell therapy generates potent and long-lasting CD4-dependent tumor immunity. <i>Sci Transl Med</i>, (2014)</li> <li>- Brown, C. E. <i>et al.</i> Optimization of IL13R<math>\alpha</math>2-Targeted Chimeric Antigen Receptor T Cells for Improved Anti-tumor Efficacy against Glioblastoma. <i>Mol Ther</i>, (2018)</li>   <li>- Dai, Z. <i>et al.</i> Ectopic CXCR2 expression cells improve the anti-tumor efficiency of CAR-T cells and remodel the immune microenvironment of pancreatic ductal adenocarcinoma. <i>Cancer Immunol Immunother</i>, (2024)</li>   <li>- Caruana, I. <i>et al.</i> Heparanase promotes tumor infiltration and antitumor activity of CAR-redirectioned T lymphocytes. <i>Nat Med</i>, (2015)</li> <li>- Wang, L.-C. S. <i>et al.</i> Targeting fibroblast activation protein in tumor stroma with chimeric antigen receptor T cells can inhibit tumor growth and augment host immunity without severe toxicity. <i>Cancer Immunol Res</i>, (2014)</li> <li>- Wehrli, M. <i>et al.</i> Mesothelin CAR T Cells Secreting Anti-FAP/Anti-CD3 Molecules Efficiently Target Pancreatic Adenocarcinoma and its Stroma. <i>Clin Cancer Res</i> <b>30</b>, 1859–1877 (2024).</li> </ul>

# Targeting Cancer Associated Fibroblast



Bughda, R. *et al. Immunotargets Ther* (2021)  
Wang, L.-C. S. *et al. Cancer Immunol Res*, (2014)



# Strategies to overcome CART resistance

Mechanism of resistance	Potential strategies to overcome	References
Unfavorable host factors		

# Strategies to overcome CART resistance

Mechanism of resistance	Potential strategies to overcome	References
Unfavorable host factors	- Pre- and post-infusion optimization of host factors	- Iacoboni, G. <i>et al.</i> Recent Bendamustine Treatment Before Apheresis Has a Negative Impact on Outcomes in Patients With Large B-Cell Lymphoma Receiving Chimeric Antigen Receptor T-Cell Therapy. <i>J Clin Oncol</i> , (2024). - Smith, M. <i>et al.</i> Gut microbiome correlates of response and toxicity following anti-CD19 CAR T cell therapy. <i>Nat Med</i> , (2022)

# Strategies to overcome CART resistance

Mechanism of resistance	Potential strategies to overcome	References
Access to CART therapy		

# Strategies to overcome CART resistance

Mechanism of resistance	Potential strategies to overcome	References
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# Strategies to overcome CART resistance

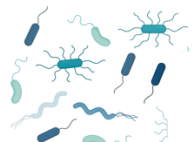
Mechanism of resistance	Potential strategies to overcome	References
Antigen loss or modulation	<ul style="list-style-type: none"> <li>- Develop CART targeting multiple antigens</li> <li>- Develop CAR with high sensitivity for the antigen</li> </ul>	<ul style="list-style-type: none"> <li>- Kokalaki, E. <i>et al.</i> Dual targeting of CD19 and CD22 against B-ALL using a novel high-sensitivity aCD22 CAR. <i>Md Ther</i>, (2023)</li> <li>- Chu, F. <i>et al.</i> Chimeric antigen receptor T cells to target CD79b in B-cell lymphomas. <i>J Immunother Cancer</i>, (2023)</li> <li>- Zhang, Y. <i>et al.</i> Safety and efficacy of a novel anti-CD19 chimeric antigen receptor T cell product targeting a membrane-proximal domain of CD19 with fast on- and off-rates against non-Hodgkin lymphoma: a first-in-human study. <i>Mol Cancer</i>, (2023)</li> </ul>
Antigen heterogeneity	<ul style="list-style-type: none"> <li>- Use CAR targeting epitopes less prone to mutation</li> </ul>	<ul style="list-style-type: none"> <li>- Cohen, J.J. <i>et al.</i> Chimeric Antigen Receptor T Cells (CART) against the IGHV4-34 B cell Receptor Eliminate Neoplastic B Cells and Reduce Antigen-Negative Escape while Sparing the 2 Healthy B cell Repertoire. Submitted</li> </ul>
CART exhaustion	<ul style="list-style-type: none"> <li>- CART gene editing for improved activation</li> <li>- Optimize co-stimulatory domains</li> </ul>	<ul style="list-style-type: none"> <li>- Guruprasad, P. <i>et al.</i> The BTLA-HVEM axis restricts CAR T cell efficacy in cancer. <i>Nat Immunol</i>, (2024)</li> <li>- Patel, R. P. <i>et al.</i> CD5 deletion enhances the antitumor activity of adoptive T cell therapies. <i>Sci Immunol</i>, (2024)</li> <li>- Doan, A. E. <i>et al.</i> FOXP1 is a master regulator of memory programming in CAR T cells. <i>Nature</i>, (2024)</li> </ul>
Lack of costimulatory signals	<ul style="list-style-type: none"> <li>- Combination with immune checkpoint blockers</li> </ul>	<ul style="list-style-type: none"> <li>- Rupp, L. J. <i>et al.</i> CRISPR/Cas9-mediated PD-1 disruption enhances anti-tumor efficacy of human chimeric antigen receptor T cells. <i>Sci Rep</i>, (2017)</li> <li>- Pérez-Moreno, M. A. <i>et al.</i> Combined or Sequential Treatment with Immune Checkpoint Inhibitors and Car-T Cell Therapies for the Management of Haematological Malignancies: A Systematic Review. <i>Int J Mol Sci</i>, (2023)</li> </ul>
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# Causes of Failure of CART Immunotherapy in the Clinic

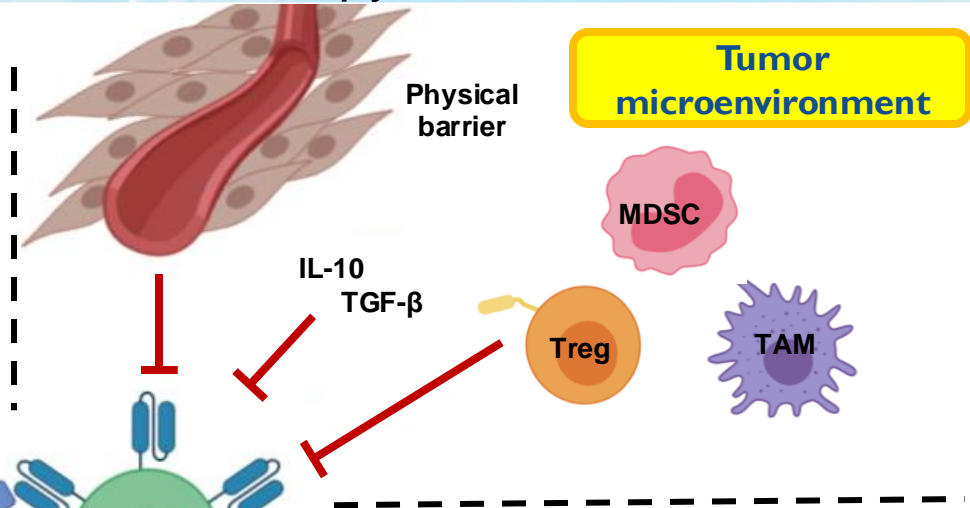
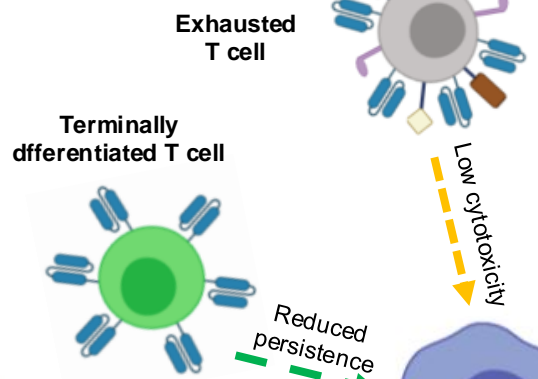
## Pre-infusion barriers

- Low lymphocyte counts
- Progression during manufacturing
- Manufacturing failure
- High Costs
- Access in general

## Host factors

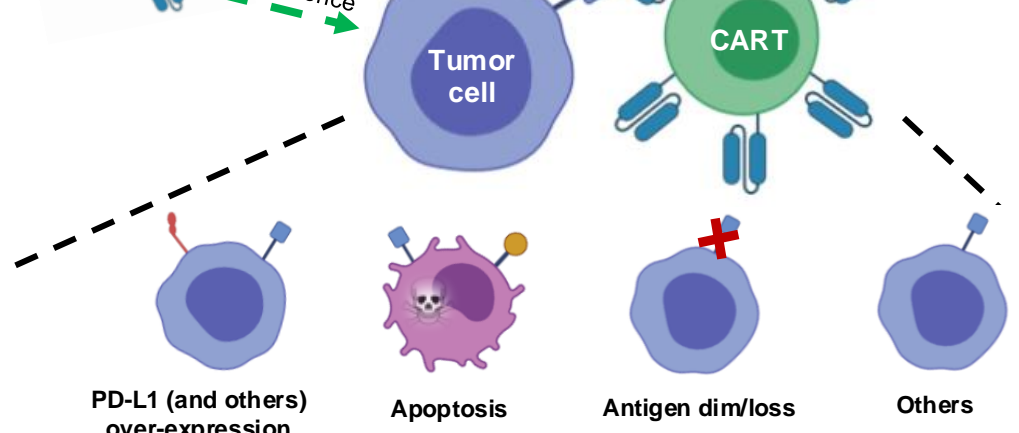
- Tumor burden, previous targeted therapies, etc
  - Immunorejection
  - Microbiota
- 

## CART dysfunction



## Tumor microenvironment

## Tumor-intrinsic mechanisms



- ICANS
- CRS
- Cytopenias
- Off-tumor
- On-target

## Toxicity

- Late neurotoxicity
- Risk of transformation

Updated from Ghilardi G., BJH, 2021

# Acknowledgements

## Ruella Lab

Patrizia Porazzi  
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Jean Lemoine  
Ruchi Patel  
Puneeth Guruprasad  
Vladlena Hornets  
Audrey Bochi-Layec  
Melody Tan  
Ekta Singh  
Ziqi Yang  
Ziyu Li  
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## Carl June Lab and CCI

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## Correlatives and manufacturing

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Stefan Barta  
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Alain Rook  
and all clinical staff

## Patients and their families

## All collaborators!!!



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# 3<sup>rd</sup> MEETING ON T-CELL AND NK-CELL BASED IMMUNOTHERAPIES FOR LYMPHOID MALIGNANCIES

Jean Lemoine, MD

**Mechanisms of resistance: laboratory evidence**

*Ruella lab, Center for Cellular Immunotherapies, University of Pennsylvania*

BOLOGNA, ROYAL HOTEL CARLTON

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